A RETRO-BIOMIMETIC ONE-STEP SYNTHESIS OF NATURAL ATISANIC AND BEYERANIC DITERPENOIDS

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Abstract: An efficient retro-biomimetic procedure for the synthesis of two natural products having atisanic and beyeranic structure is reported. The readily available *ent*-kaurenoic acid is used as the substrate, which provides under superacidic treatment a mixture of natural *ent*-beyer-15-en-19-oic and *ent*-atis-16-en-19-oic acids. These compounds have been reported as components of medicinal plants and possess relevant biological activity. Their structure was confirmed on the basis of chemical transformations and spectral data.

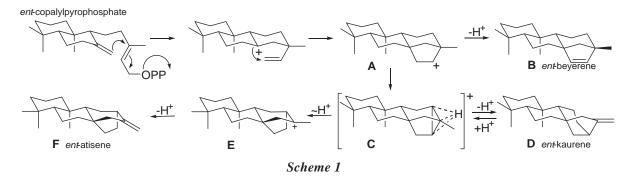
Keywords: Terpenoids, biomimetic synthesis, kaurane, atisane, beyerane.

1. Introduction

Diterpenoids represent a large family of natural products with a broad spectrum of biological activities. Continuous work on diterpenes reveal more and more representatives of this large group to possess interesting properties. Examples are the well known atisanic and beyeranic diterpenoids. They have been known since quite a long time, but their biological activity remained still unexplored. Only recent studies revealed quite surprising data. For example the *ent*-beyer-15-en-19-oic acid **1** isolated from the roots of the Mexican medicinal plant *Viguiera hypargyrea* showed antispasmodic and antimicrobial activity in low concentrations [1]. The related *ent*-atis-16-en-19-oic acids **2** was shown recently to be a oviposition stimulant for the banded sunflower moth, *Cochylis hospes* [2]. We present in the current communication our first results on the synthesis of natural acids **1** and **2**, basing on a one-step retro-biomimetic strategy.

2. Results and discussion

The beyeranic and atisanic diterpenoids posess a complex structure of condensed and bridged cyclic systems. Due to the complexity of these skeletons, total synthesis of these compounds proved to be quite difficult. In fact, there are only several publications relating on the total synthesis of beyeranes or atisanes. The synthetic strategies include multiple steps and the overall performance is relatively low [3, 4]. On the other hand, partial synthesis of atisanes and beyeranes have been reported, basing on biomimetic-like transformations of other available natural products like trachylobanic acid [5] or *iso*-steviol [6, 7]. These approaches proved to be more efficient, both from the point of view of length and overall yield. Basing on this conclusion, we have developed a semi-synthetic approach for the synthesis of above mentioned acids **1** and **2** starting from readily available [8] *ent*-kaurenoic acid **3**. This approach is very simple, and involves a single step transformation of the substrate to the target molecules.



The transformation is a biomimetic, acid induced rearrangement of the *ent*-kauranic framework to atisane and beyerane compounds. More exactly, this is a retro-biomimetic procedure, since the beyeranic framework is regarded as biogenetical precursor for *ent*-kauranic compounds. This hypothesis [9] has been formulated on the basis of the known isoprenic rule and to the best of our knowledge was not turned down yet. Scheme 1 provides a general overview of this biogenetical scheme, and our primarily intention was to perform the retro-biomimetic transformation of kaurene D to beyerene B or atisene F (path D->C---B or F).

The rearrangements of *ent*-kauranic diterpenes has been reported under the action of different reagents [10]. Most of the examples relate on the reactions involving the formation of the non-classical carbocation of type C (Scheme 1). It is well known from the work of Olah, that superacids are better generators of these species and

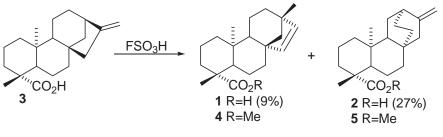
our own experience on the use of fluorosulfonic acid as an efficient promoter of terpenoids cyclization has provided a motivation to investigate the behavior of *ent*-kaurenic substrates under superacidic treatment. The mild reaction conditions (-78°C), as well as a moderate excess of the superacid can contribute to a sufficient lifetime of generated carbonium ions so that to assure the rearrangement of *ent*-kaurene skeleton in a more selective manner.

Submission of the substrate **3** to this reaction condition [12] has shown that isomerisation takes place smoothly, providing a basic product, having similar chromatographical behavior as starting material. Only a small fraction of more polar byproducts was detected. The reaction pathway proved to be not sensitive to reaction conditions variation (temperature, duration, quantity of superacid).

The basic reaction product, although chromatographically homogeneous, showed a complex NMR spectrum. Analysis of silver nitrate impregnated TLC plates revealed the presence of at least 3 components, which were tentatively separated by argentation column chromatography. One pure individual compounds was isolated and its structure was determined by extensive NMR studies. The determined structure corresponded to a atisane skeleton with the exocyclic double bond 2 [13].

The mixture of acids that was considered unresolved was further separated by semi-preparative HPLC, using a normal phase column. The major isolated compound was the starting *ent*-kaurenic acid 3 (19%). A minor amount (9%, on the basis of recovered 3) of beyerane 1 [14] was also obtained. Both 1 and 2 were methylated with an ethereal solution of diazomethane, to provide individual methyl esters 4 [15] and 5 [16] respectively (Scheme 2). The analytical data of the acids 1 and 2, as well as of the corresponding esters 5 and 6 matched perfectly the published data.

A minor fraction of the reaction product (cca. 30%) was eluted in the HPLC experiment unresolved. The possibility of isolation from this fraction of individual isomerisation compounds are currently under investigation.



Scheme 2

3. Conclusions

The direct, one step conversion of *ent*-kaurenoic acid **3** into atisanic and beyeranic acids **1** and **2** was performed under the action of fluorosulfonic acid.

4. Acknowledgements

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5. References and notes

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- [12]. A solution of *ent*-kaurenoic acid 3 (403 mg, 1.324 mmol) in a mixture of *i*-PrNO₂ (2 ml) and DCM (9 ml), cooled at -70 °C, was treated with FSO₃H (662 mg, 6.623 mmol) in *i*-PrNO₂ (1.4 ml), under stirring. After 15 min., the reaction was stopped by adding a solution of Et₂N (4 ml) in light petroleum ether (4 ml). The usual work up gave

415 mg of a crude residue, which was submited to flash chromatography. Elution with a mixture of EtOAc in petroleum ether (2%) gave 298 mg (74%) of a TLC-homogenious reaction product.

- [13]. *ent*-Atis-16-en-19-oic acid **2**. IR (neat): 2918, 1689, 1464, 1446, 1273, 1258 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): $\delta = 4.73$ (1H, bd, J=2Hz), 4.57 (1H, bd, J=2Hz), 2.22 (1H, bs), 2.15 (1H, bd, J=14Hz), 2.04 (1H, bd, J=17Hz), 1.29-2.00 (m, 19H), 1.25 (3H, s), 1.24 (3H, s), 0.91-1.18 (m, 8H), 0.90 (3H, s), 0.70-0.88 (m, 8H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 182.33$ (s, C-19), 152.77 (s, C-15), 104.52 (t, C-17), 57.17 (d, C-5), 52.21 (d, C-9), 48.21 (t, C-16), 43.70 (s, C-4), 39.73 (t, C-1), 39.66 (t, C-7), 38.40 (s, C-10), 38.08 (t, C-3), 36.61 (d, C-12), 33.55 (s, C-8), 28.90 (q, C-18), 28.73 (t, C-14), 28.32 (t, C-13), 27.27 (t, C-11), 20.27 (t, C-6), 18.76 (t, C-2), 12.10 (q, C-20).
- [14] *ent*-Beyer-15-en-19-oic acid **1**. IR (neat): 2935, 1685, 1445, 1255, 1190 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): $\delta = 5.76$ (1H, d, J=5.7Hz), 5.47 (1H, d, J=5.7 Hz), 1.26 (3H, s), 1.01 (3H, s), 0.69 (3H, s). ¹³C NMR (100 MHz, CDCl₃): $\delta = 184.3$ (s, C-19), 136.5 (d, C-16), 134.8 (d, C-15), 61.1 (t, C-14), 57.2 (d, C-5), 52.4 (d, C-9), 49.2 (s, C-8), 43.9 (s, C-4), 43.7 (s, C-13), 39.6 (t, C-1), 38.0 (t, C-3), 37.7 (t, C-7), 37.7 (s, C-10), 33.2 (t, C-12), 29.1 (q, C-18), 24.9 (q, C-17), 21.6 (t, C-6), 20.5 (t, C-11), 19.3 (t, C-2), 13.8 (q, C-20).
- [15]. Methyl-ent-beyer-15-en-19-oate 4. ¹H NMR (400 MHz, CDCl₃): δ = 5.74 (1H, d, J=5.7Hz), 5.48 (1H, d, J=5.7 Hz), 3.60 (3H, s), 1.20 (3H, s), 1.02 (3H, s), 0.59 (3H, s). ¹³C NMR (100 MHz, CDCl₃): δ = 178.08 (s, C-19), 134.77 (d, C-16), 136.52 (d, C-15), 61.09 (t, C-14), 57.15 (d, C-5), 51.09 (q, -CO₂Me), 52.32 (d, C-9), 49.16 (s, C-8), 43.66 (s, C-4), 43.66 (s, C-13), 39.59 (t, C-1), 38.23 (t, C-3), 37.67 (t, C-7), 37.76 (s, C-10), 33.15 (t, C-12), 28.97 (q, C-18), 24.87 (q, C-17), 21.67 (t, C-6), 20.46 (t, C-11), 19.33 (t, C-2), 13.64 (q, C-20).
- [16]. Methyl-*ent*-atis-16-en-19-oate **5**. ¹H NMR (400 MHz, CDCl₃): $\delta = 4.74$ (1H, bd, J=2Hz), 4.58 (1H, bd, J=2Hz), 3.66 (3H, s), 1.17 (3H, s), 0.81 (3H, s). ¹³C NMR (100 MHz, CDCl₃): $\delta = 177.94$ (s, C-19), 152.77 (s, C-15), 104.50 (t, C-17), 57.25 (d, C-5), 52.19 (d, C-9), 51.03 (q, -CO₂<u>Me</u>), 48.23 (t, C-16), 43.90 (s, C-4), 39.70 (t, C-1), 39.70 (t, C-7), 38.20 (s, C-10), 38.28 (t, C-3), 36.63 (d, C-12), 33.54 (s, C-8), 28.73 (q, C-18), 28.72 (t, C-14), 28.31 (t, C-13), 27.27 (t, C-11), 20.34 (t, C-6), 18.84 (t, C-2), 11.95 (q, C-20).